

Nat. Prod. Bioprospect. (2014) 4:47–51
DOI 10.1007/s13659-014-0003-9

ORIGINAL ARTICLE

New Taxane Diterpenoids from *Taxus yunnanensis*

Ping Hai · Shi-Zhen Wen · Yan Li ·
Yuan Gao · Xian-Jun Jiang · Fei Wang



Received: 26 November 2013 / Accepted: 12 January 2014 / Published online: 19 February 2014
© The Author(s) 2014. This article is published with open access at Springerlink.com

Abstract Three hitherto unknown taxane diterpenoids, namely baccatin VIII (**1**), baccatin IX (**2**), and baccatin X (**3**), along with 10 known analogues were isolated from an ethanolic extract of the twigs and leaves of *Taxus yunnanensis*. The new structures were characterized based on extensive spectroscopic analysis. Compounds **1** and **2** were tested for their in vitro cytotoxicity against five human tumor cell lines, and **1** exhibited inhibitory effects on HL-60 and MCF-7, with IC₅₀ values of 3.44 and 9.67 μM, respectively.

Keywords *Taxus yunnanensis* · Taxane · Baccatin · Cytotoxicity

1 Introduction

Plants of the genus *Taxus* (Taxaceae) are large, ornamental evergreen shrubs or trees, most of which are distributed in the Northern hemisphere and have attracted much attention owing to natural taxane diterpenoid Taxol, an antitumor agent for treating the sufferers of ovarian, breast, and other carcinomas [1, 2]. Several groups conducted a series of phytochemical research work on the *Taxus* species, and more than 550 taxane diterpenoids have been isolated from this genus [2–5]. *Taxus yunnanensis*, a slow-growing tree commonly known as “Yunnan Hongdoushan” and grown mainly in Yunnan Province of China, is considered as a promising source of taxane diterpenes [6]. With the aim of

isolating more Taxol derivatives with biological activities, we investigated the constituents of the twigs and leaves of *T. yunnanensis* once again, which led to the isolation of three new taxane diterpenoids baccatins VIII, IX, and X (**1–3**), together with 10 known analogues, baccatin III (**4**) [7], 10-deacetyl baccatin III (**5**) [7], 19-hydroxybaccatin III (**6**) [8], 14β-benzoyloxy-2-deacetyl baccatin VI (**7**) [9], baccatin IV (**8**) [10], 1-hydroxybaccatin I (**9**) [7], 13-*O*-deacetyl taxumairol Z (**10**) [11], taxayunnansin A (**11**) [12], 2-deacetoxytaxinine B (**12**) [13], and taxinine B (**13**) [14]. This paper reports on the isolation and structure determination of the new taxanes and their cytotoxicity.

2 Results and Discussion

Compound **1** was obtained as a white amorphous powder. The HRESIMS data (m/z 669.2513 [M + Na]⁺) indicated the molecular formula C₃₃H₄₂O₁₃, requiring 13 degrees of unsaturation. The IR absorption bands at 3433 and 1726 cm^{−1} suggested the presence of hydroxy and carbonyl functionalities, respectively. The ¹H NMR spectrum of **1** (Table 1) displayed signals of four tertiary methyls (δ_H 1.19, 1.65, 1.77, and 1.88), three acetyl methyls (δ_H 2.10, 2.23, and 2.34), seven oxygenated methines (δ_H 4.03, 4.37,

Electronic supplementary material The online version of this article (doi:10.1007/s13659-014-0003-9) contains supplementary material, which is available to authorized users.

P. Hai · S.-Z. Wen · Y. Gao · X.-J. Jiang · F. Wang (✉)
BioBioPha Co., Ltd., Kunming 650201, China
e-mail: wangfei@mail.kib.ac.cn

Y. Li · Y. Gao · F. Wang
State Key Laboratory of Phytochemistry and Plant Resources in
West China, Kunming Institute of Botany, Chinese Academy of
Sciences, Kunming 650201, China

Table 1 NMR data for compounds **1–3**

| No. | 1 ^a | | 2 ^b | | 3 ^a | |
|----------------------|---------------------------------------|---------------------|---------------------------------------|---------------------|---------------------------------------|---------------------|
| | δ_{H} (<i>J</i> in Hz) | δ_{C} | δ_{H} (<i>J</i> in Hz) | δ_{C} | δ_{H} (<i>J</i> in Hz) | δ_{C} |
| 1 | | 76.8 (s) | | 75.6 (s) | | 79.0 (s) |
| 2 | 5.87 (d, 6.1) | 73.8 (d) | 5.65 (d, 6.0) | 72.7 (d) | 4.02 (d, 6.0) | 74.2 (d) |
| 3 | 3.01 (d, 6.1) | 47.1 (d) | 2.89 (d, 6.0) | 46.0 (d) | 2.86 (d, 6.0) | 48.7 (d) |
| 4 | | 82.8 (s) | | 80.8 (s) | | 83.9 (s) |
| 5 | 4.92 (br. d, 9.0) | 85.2 (d) | 4.82 (br. d, 9.2) | 83.3 (d) | 5.00 (dd, 9.3, 1.3) | 85.2 (d) |
| 6 α | 2.46 (ddd, 15.0, 9.0, 8.0) | 38.2 (t) | 2.28 (ddd, 14.8, 9.2, 7.2) | 37.7 (t) | 2.47 (ddd, 14.7, 9.3, 7.5) | 38.8 (t) |
| 6 β | 1.84 (ddd, 15.0, 10.0, 0.9) | | 1.63 (ddd, 14.8, 10.3, 0.8) | | 1.86 (ddd, 14.7, 10.0, 1.3) | |
| 7 | 4.37 (dd, 10.0, 8.0) | 74.5 (d) | 4.21 (overlap) | 73.2 (d) | 4.41 (dd, 10.0, 7.5) | 75.4 (d) |
| 8 | | 45.4 (s) | | 44.0 (s) | | 45.6 (s) |
| 9 | 4.51 (d, 11.0) | 77.6 (d) | 4.25 (overlap) | 76.1 (d) | 4.29 (d, 11.0) | 78.3 (d) |
| 10 | 6.17 (d, 11.0) | 74.2 (d) | 6.01 (d, 10.8) | 73.3 (d) | 6.21 (d, 11.0) | 75.0 (d) |
| 11 | | 137.7 (s) | | 134.3 (s) | | 136.4 (s) |
| 12 | | 137.3 (s) | | 141.1 (s) | | 142.3 (s) |
| 13 | 6.00 (dq, 6.7, 0.9) | 79.8 (d) | 4.38 (br. dd, 5.9, 5.6) | 75.5 (d) | 4.94 (overlap) | 74.3 (d) |
| 14 | 4.03 (d, 6.7) | 70.5 (d) | 3.80 (dd, 6.6, 5.9) | 72.4 (d) | 5.05 (d, 6.1) | 79.1 (d) |
| 15 | | 43.8 (s) | | 42.2 (s) | | 43.8 (s) |
| 16 | 1.19 (s) | 28.7 (q) | 0.96 (s) | 28.4 (q) | 1.29 (s) | 28.9 (q) |
| 17 | 1.65 (s) | 24.4 (q) | 1.46 (s) | 23.9 (q) | 1.52 (s) | 23.8 (q) |
| 18 | 1.88 (d, 0.9) | 14.9 (q) | 1.90 (s) | 15.2 (q) | 2.06 (d, 1.3) | 15.6 (q) |
| 19 | 1.77 (s) | 13.0 (q) | 1.60 (s) | 12.5 (q) | 1.77 (s) | 13.1 (q) |
| 20 α | 4.21 (d, 8.2) | 77.3 (t) | 3.96 (br. s) | 75.4 (t) | 4.70 (d, 8.8) | 79.1 (t) |
| 20 β | 4.17 (d, 8.2) | | | | 4.60 (d, 8.8) | |
| 1' | | 167.1 (s) | | 165.4 (s) | | 169.3 (s) |
| 2' | | 130.6 (s) | | 129.9 (s) | | 131.4 (s) |
| 3', 7' | 8.07 (br. d, 7.8) | 130.8 (d) | 8.02 (br. d, 7.8) | 129.9 (d) | 8.09 (br. d, 8.3) | 130.9 (d) |
| 4', 6' | 7.48 (dd, 7.8, 7.6) | 129.4 (d) | 7.51 (dd, 7.8, 7.6) | 128.8 (d) | 7.49 (dd, 8.3, 7.6) | 129.5 (d) |
| 5' | 7.61 (br. t, 7.6) | 134.3 (d) | 7.64 (br. t, 7.6) | 133.4 (d) | 7.62 (br. t, 7.6) | 134.4 (d) |
| 4-COCH ₃ | 2.34 (s) | 22.9 (q) | 2.18 (s) | 22.7 (q) | 2.08 (s) | 22.9 (q) |
| 4-COCH ₃ | | 171.2 (s) | | 169.7 (s) | | 172.0 (s) |
| 10-COCH ₃ | 2.10 (s) | 21.3 (q) | 2.01 (s) | 21.2 (q) | 2.08 (s) | 21.3 (q) |
| 10-COCH ₃ | | 171.9 (s) | | 169.9 (s) | | 172.2 (s) |
| 13-COCH ₃ | 2.23 (s) | 21.2 (q) | | | | |
| 13-COCH ₃ | | 172.3 (s) | | | | |
| 1-OH | | | 4.40 (s) | | | |
| 7-OH | | | 6.30 (d, 3.5) | | | |
| 9-OH | | | 6.31 (d, 3.0) | | | |
| 13-OH | | | 5.46 (d, 5.6) | | | |
| 14-OH | | | 6.74 (d, 6.6) | | | |

^a Measured in CD₃OD^b Measured in DMSO-*d*₆

4.51, 4.92, 5.87, 6.00, and 6.17), a benzoyl group (δ_{H} 8.07 \times 2, 7.48 \times 2, and 7.61), and an oxetane ring (δ_{H} 4.21 and 4.17, each 1H, d, *J* = 8.2 Hz). Beside the resonances for three acetyl groups (δ_{C} 171.2, 22.9; 171.9, 21.3; 172.3, 21.2) and a benzoyl group (δ_{C} 167.1, 130.6,

129.4 \times 2, 130.8 \times 2, and 134.3), 20 carbon signals, including an oxygenated methylene carbon (δ_{C} 77.3), seven oxygenated methines (δ_{C} 70.5, 73.8, 74.2, 74.5, 77.6, 79.8, 85.2), two oxygenated quaternary carbons (δ_{C} 76.8 and 82.8), and a tetrasubstituted double bond (δ_{C} 137.3 and

137.7) were displayed in the ^{13}C NMR and DEPT spectra of **1** (Table 1). Comparison of the NMR data with those of 9-dihydro-13-*O*-acetylbaccatin III [15] revealed that **1** was a taxane diterpenoid very similar to it. The only difference between them was that a methylene (δ_{C} 36.1, C-14) in 9-dihydro-13-*O*-acetylbaccatin III was replaced by a hydroxylated methine (δ_{C} 70.5) in **1**. This was further confirmed by the HMBC cross-peaks of H-14 (δ_{H} 4.03, d, $J = 6.7$ Hz) with C-1 (δ_{C} 76.8, s), C-13 (δ_{C} 79.8, d), and C-15 (δ_{C} 43.8, s).

The relative configuration of **1** was deduced from the ROESY experiment (Fig. 1). The significant correlations of H-14/H-3 and H-3/H-7 were observed, indicating α -orientation of these protons. Similarly, the correlations of H-2/Me-19, Me-19/H-9, and Me-19/H-20 β were observed, revealing that these protons were on the other face of the molecule. β -Orientation of H-13 was suggested by the strong correlation between H-13 and Me-16, while α -

orientation of H-10 and H-5 was deduced by the correlations of H-10/Me-18 and H-5/H-20 α , respectively. In addition, the hydroxy group attached to the bridgehead carbon was assigned as β -oriented, based on ring junction inferred by the correlations of H-13/Me-16 and H-14/H-3. Thus, the structure of **1** was unambiguously identified as shown, and given the trivial name baccatin VIII.

Compound **2** was isolated as a white powder with the molecular formula $\text{C}_{31}\text{H}_{40}\text{O}_{12}$, as determined by HREIMS: m/z 604.2515 $[\text{M}]^+$ (calcd for $\text{C}_{31}\text{H}_{40}\text{O}_{12}$, 604.2520). Analysis of its NMR data (Table 1) indicated that **2** closely resembled **1** except for the absence of signals for an acetyl and a dramatic upfield shift of H-13 from δ_{H} 6.00 in **1** to δ_{H} 4.38 in **2**. This indicated an absence of the acetyl at C-13, as established by the HMBC correlations of 13-OH (δ_{H} 5.46, d, $J = 5.6$ Hz) with C-12 (δ_{C} 141.1), C-13 (δ_{C} 75.5), and C-14 (δ_{C} 72.4). The relative configuration of 13-OH was assigned as α on the basis of the ROESY correlation of H-13 with Me-16. The stereochemistry of other chiral centers was in accordance with that of **1**, by the ROESY correlations of H-2/Me-19, H-3/H-7, H-3/H-5, H-3/H-14, H-10/Me-18, and Me-19/H-20 β . The structure of **2** was therefore characterized as shown and named baccatin IX.

Compound **3** gave the same molecular formula ($\text{C}_{31}\text{H}_{40}\text{O}_{12}$) as **2** by negative HRESIMS: m/z 603.2445 $[\text{M} - \text{H}]^-$ (calcd for $\text{C}_{31}\text{H}_{39}\text{O}_{12}$, 603.2442). Comparison of the NMR data (Table 1) of **3** with those of **2** implied that they were very similar. However, the HMBC correlations of H-2 (δ_{H} 4.02, d, $J = 6.0$ Hz) with C-1 (δ_{C} 79.0), C-3 (δ_{C} 48.7), C-8 (δ_{C} 45.6), and C-15 (δ_{C} 43.8), and H-14 (δ_{H} 5.05, d, $J = 6.1$ Hz) with C-13 (δ_{C} 74.3) and C-15 (δ_{C} 43.8) revealed that H-2 was shifted upfield from δ_{H} 5.65 in **2** to δ_{H} 4.02 in **3**, while H-14 was shifted downfield from δ_{H} 3.80 in **2** to δ_{H} 5.05 in **3**. The above two remarkable shifts ($\Delta - 1.63$ ppm of H-2; $\Delta + 1.25$ ppm of H-14) implied the deesterification and esterification related to aromatic ester, which required a transfer of the benzoyl group from C-2 in **2** to C-14 in **3**. This deduction was further verified by the HMBC correlation of H-14 with the benzoyl carbonyl carbon at δ_{C} 169.3. The relative configuration of **3** was established as shown by comparison of relevant coupling constants with those of the isolated analogues. Accordingly, the structure of **3** was determined and named baccatin X.

Compounds **1** and **2** were tested for their cytotoxicity in vitro against five human cancer cell lines: HL-60, SMMC-7721, A-549, MCF-7, and SW480. As summarized in Table 2, **1** was cytotoxic for all the cell lines tested, and displayed moderate activities against HL-60 and MCF-7 (IC_{50} 3.44 and 9.67 μM , respectively), while **2** showed selective cytotoxicity against some of the cell lines.

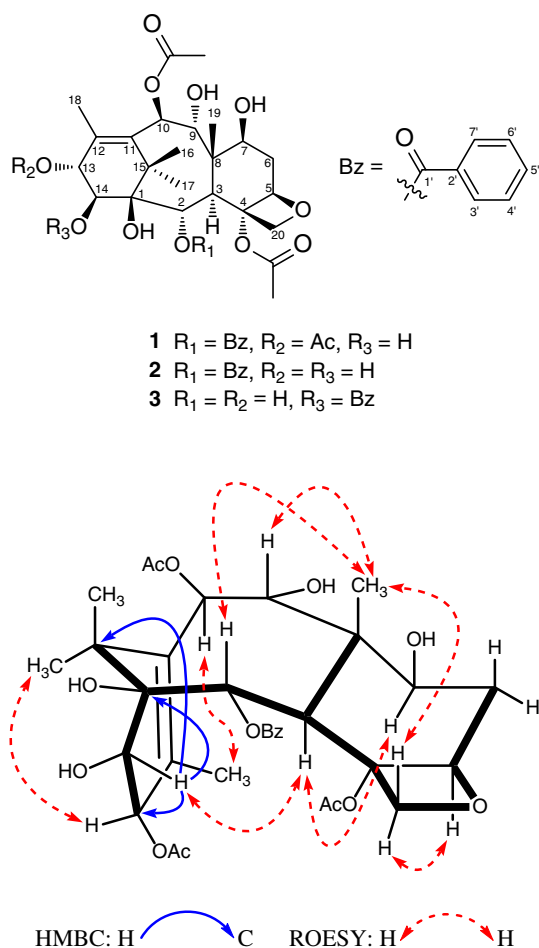


Fig. 1 Key HMBC and ROESY correlations of **1**

Table 2 Cytotoxicity data for **1** and **2** with IC₅₀ values (μM)

| Compound | HL-60 | SMMC-7721 | A-549 | MCF-7 | SW480 |
|------------------------|--------|-----------|--------|--------|--------|
| 1 | 3.44 | 14.83 | 21.27 | 9.67 | 22.42 |
| 2 | 20.23 | 16.69 | >40 | 25.32 | >40 |
| Cisplatin ^a | 1.06 | 4.32 | 5.08 | 15.41 | 15.35 |
| Taxol ^a | <0.008 | <0.008 | <0.008 | <0.008 | <0.008 |

^a Positive controls

3 Experimental Section

3.1 General Experimental Procedures

Optical rotations were measured in methanolic solution on a Jasco P-1020 automatic digital polarimeter. IR spectra in potassium bromide discs were taken with a Bruker Tensor 27 FT-IR spectrometer. NMR spectra were performed on Bruker DRX-500 and Avance III 600 instruments with deuterated solvent signals as internal standards. MS data were measured on VG Auto Spec-3000, API QSTAR time-of-flight, and Bruker Esquire HCT spectrometers. Column chromatography was carried out with silica gel (200–300 mesh) and Sephadex LH-20 (Amersham Biosciences, Sweden). Fractions were monitored by TLC and reversed-phase HPLC (Agilent 1200, Agilent Zorbax Extend-C18 column, 5 μm, 4.6 × 150 mm).

3.2 Plant Material

The twigs and leaves of *T. yunnanensis* were collected from Yunnan Province of China in February 2008, and identified by Mr. Yu Chen of Kunming Institute of Botany, Chinese Academy of Sciences. A voucher specimen (No. BBP0026016TY) was deposited at BioBioPha Co., Ltd.

3.3 Extraction and Isolation

Dried and powdered twigs and leaves (7.0 kg) of *T. yunnanensis* were extracted with EtOH-H₂O (95:5, v/v; 3 × 12 L, each 5 days) at room temperature. The combined filtrate was concentrated under vacuum and fractionated by silica gel CC successively eluted with a gradient of increasing acetone in petroleum ether (PE) to obtain four fractions (A–D). Fraction B (PE/Me₂CO = 6:1) was first chromatographed on silica gel columns, and each major fraction was then purified using a Sephadex LH-20 column (CHCl₃/MeOH = 1:1) to yield compounds **8** (8 mg), **12** (11 mg), and **13** (45 mg). Fraction C (PE/Me₂CO = 3:1) was subjected to silica gel CC, eluting with CHCl₃/MeOH (100:1 → 0:100) to give four fractions (C1–C4). After repeated CC on silica gel (CHCl₃/Me₂CO, 10:1 → 0:1) and Sephadex LH-20 (CHCl₃/MeOH = 1:1), fraction C4 gave compounds **1** (64 mg), **2** (47 mg), **3** (2 mg),

5 (192 mg), **6** (243 mg), and **10** (66 mg). Fraction D (PE/Me₂CO = 1:1) was purified by repeated CC over silica gel (CHCl₃/MeOH, 100:1 → 0:100) and recrystallization to afford compounds **4** (327 mg), **7** (226 mg), **9** (884 mg), and **11** (264 mg).

3.4 Baccatin VIII (**1**)

White powder, $[\alpha]_D^{27} -15.4$ (c 0.13, MeOH); UV (MeOH) λ_{\max} : 230, 274 nm; IR (KBr) ν_{\max} : 3433, 2994, 2896, 1726, 1629, 1439, 1373, 1239, 1110, 1054, 1025, 757, 713 cm⁻¹; ¹H and ¹³C NMR data: see Table 1; ESIMS (pos.): *m/z* 669 [M + Na]⁺; HRESIMS (pos.): *m/z* 669.2513 [M + Na]⁺ (calcd for C₃₃H₄₂O₁₃Na, 669.2523).

3.5 Baccatin IX (**2**)

White powder, $[\alpha]_D^{20} +17.3$ (c 0.18, MeOH); UV (MeOH) λ_{\max} : 230, 274 nm; IR (KBr) ν_{\max} : 3428, 1739, 1719, 1633, 1452, 1435, 1372, 1273, 1168, 1071, 1026, 985, 946, 714 cm⁻¹; ¹H and ¹³C NMR data: see Table 1; ESIMS (pos.): *m/z* 627 [M + Na]⁺; HREIMS: *m/z* 604.2515 (calcd for C₃₁H₄₀O₁₂, 604.2520).

3.6 Baccatin X (**3**)

White powder; UV (MeOH) λ_{\max} : 230, 273 nm; ¹H and ¹³C NMR data: see Table 1; ESIMS (pos.): *m/z* 627 [M + Na]⁺; HRESIMS (neg.): *m/z* 603.2445 [M – H][–] (calcd for C₃₁H₃₉O₁₂, 603.2442).

3.7 Cytotoxicity Bioassays

The following human tumor cell lines were used: HL-60, SMMC-7721, A-549, MCF-7, and SW480. All cells were cultured in RPMI-1640 or DMEM medium (Hyclone, Logan, UT, USA), supplemented with 10 % fetal bovine serum (Hyclone) at 37 °C in a humidified atmosphere with 5 % CO₂. The cytotoxicity assay was performed using the modified MTS (3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium) method as previously described [16]. The IC₅₀ value of each compound was calculated by the Reed and Muench's method [17].

Acknowledgments This work was financially supported by the “Large-scale Compound Library” project of National Development and Reform Commission of China and the “Western Light” talents training program of Chinese Academy of Sciences.

Open Access This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

References

1. E. Baloglu, D.G.I. Kingston, *J. Nat. Prod.* **62**, 1448 (1999)
2. Y.F. Wang, Q.W. Shi, M. Dong, H. Kiyota, Y.C. Gu, B. Cong, *Chem. Rev.* **111**, 7652 (2011)
3. Q. Yue, Q.C. Fang, X.T. Liang, C.H. He, *Phytochemistry* **39**, 871 (1995)
4. L.O. Zamir, J.Z. Zhang, J.H. Wu, F. Sauriol, O. Mamer, *Tetrahedron* **55**, 14323 (1999)
5. Q. Yue, Q.C. Fang, X.T. Liang, *Phytochemistry* **43**, 639 (1996)
6. N.T. Nguyen, A.H. Banskota, Y. Tezuka, T. Nobukawa, S. Kadota, *Phytochemistry* **64**, 1141 (2003)
7. R.W. Miller, R.G. Powell, C.R. Smith Jr, E. Arnold, J. Clardy, *J. Org. Chem.* **46**, 1469 (1981)
8. J.L. McLaughlin, R.W. Miller, R.G. Powell, C.R. Smith Jr, *J. Nat. Prod.* **44**, 312 (1981)
9. F.S. Wang, L.Y. Peng, Y. Zhao, G. Xu, Q.S. Zhao, H.D. Sun, *J. Nat. Prod.* **67**, 905 (2004)
10. F.S. Wang, L.Y. Peng, Y. Zhao, K. Gu, Q.S. Zhao, H.D. Sun, *Acta Bot. Yunnan.* **25**, 369 (2003)
11. Y.C. Shen, Y.T. Chang, S.S. Wang, Y.C. Lin, C.Y. Chen, *Chem. Pharm. Bull.* **50**, 1561 (2002)
12. H.J. Zhang, Y. Takeda, H.D. Sun, *Phytochemistry* **39**, 1147 (1995)
13. J.Y. Liang, K.S. Huang, A.A.L. Gunatilaka, L. Yang, *Phytochemistry* **47**, 69 (1998)
14. Y. Konda, T. Sasaki, X.L. Sun, X. Li, M. Onda, H. Takayanagi, Y. Harigaya, *Chem. Pharm. Bull.* **42**, 2621 (1994)
15. G.P. Gunawardana, U. Premachandran, N.S. Burres, D.N. Whittern, R. Henry, S. Spanton, J.B. McAlpine, *J. Nat. Prod.* **55**, 1686 (1992)
16. Y. Gao, G.T. Li, Y. Li, P. Hai, F. Wang, J.K. Liu, *Nat. Prod. Bioprospect.* **3**, 14 (2013)
17. L.J. Reed, H. Muench, *Am. J. Hyg.* **27**, 493 (1938)